# The comparisons of blood plasma and cerebrospinal fluid S100B protein concentrations in patients with Alzheimer's disease, amyotrophic lateral sclerosis, and multiple sclerosis

Tarasiuk J.  $^{1,A-F}$ , Kapica-Topczewska K.  $^{1,A,B,D,F}$ , Chorąży M.  $^{1,C}$ , Borowik-Zaręba A.  $^{2,A,C}$ , Mroczko B.  $^{3,C,E}$ , Kochanowicz J.  $^{1,E}$ , Kułakowska A.  $^{1,A-F}$ 

- 1. Department of Neurology, Medical University of Bialystok, Bialystok, Poland.
- 2. Department of Neurology, Wojewódzki Szpital Zespolony im. Jędrzeja Śniadeckiego w Białymstoku, Białystok, Poland.
- 3. Department of Neurodegeneration Diagnostics, Medical University of Bialystok, Bialystok, Poland.

**A**- Conception and study design; **B** - Collection of data; **C** - Data analysis; **D** - Writing the paper;

E- Review article; F - Approval of the final version of the article; G - Other (please specify)

# ABSTRACT

**Introduction:** S100 calcium-binding protein B (S100B) is a biochemical marker of astroglial damage.

**Purpose:** To assess the pathophysiological implications of S100B concentrations in blood plasma and cerebrospinal fluid of patients with neurodegenerative central nervous system disorders. **Materials and Methods:** In this study, we determined and compare S100B concentrations in blood plasma and cerebrospinal fluid obtained from subjects diagnosed with Alzheimer's disease (n=20), amyotrophic lateral sclerosis (n=12), multiple sclerosis (n=40) and the reference group (n=20), using enzyme-linked immunosorbent assay.

**Results:** Concentrations of S100B in plasma collected from patients diagnosed with Alzheimer's disease (252,38±183,50 pg/mL) and multiple sclerosis (164,92±250,14 pg/mL) were above laboratory standards, but in patients with amyotrophic lateral sclerosis (53,96±56,92 pg/mL) and the reference group (2,12 pg/mL) were below

laboratory norms (N>75 pg/mL). Concentrations of S100B in plasma collected from patients with Alzheimer's disease (252,38±183,50 pg/mL) were significantly higher than in patients with amyotrophic lateral sclerosis (53,96±56,92 pg/mL) (p<0,029). Concentrations of S100B in CSF collected from the reference group (546,96±236,62 pg/mL) and from patients with Alzheimer's disease (587,53±189,57 pg/mL), amyotrophic lateral sclerosis (404,41±179,56 pg/mL), multiple sclerosis (462,03±146,01 pg/mL) were very similar, and none of pairwise comparisons reached statistical significance.

**Conclusions:** Results of our studies indicate the importance of S100B protein concentration assessment in blood in central nervous system disorders differential diagnostics.

**Keywords**: S100, blood, cerebrospinal fluid, Alzheimer's disease, Amyotrophic lateral sclerosis, Multiple sclerosis

# DOI:

\*Corresponding author:

Department of Neurology, Medical University of Bialystok, Sklodowskiej-Curie 24 A, 15-276 Bialystok, Poland phone: +48 85 746 8427, Fax: +48 85 746 86 08; e-mail: amirtarasiuk@wp.pl

Received: 15.01. 2019 Accepted: 12.03.2019 Progress in Health Sciences Vol. 9(1) 2019 pp 22-27

© Medical University of Białystok, Poland

# **INTRODUCTION**

S100 calcium-binding protein B (S100B) is a member of the S100 protein family, synthesized in astrocytes and Schwann cells, adipocytes, chondrocytes, lymphocytes, bone marrow and melanoma cells [1]. The gene encoding S100B is located at 21q22.3 chromosome. Intracellularly, S100B is a normal part of calcium hemostasis, thereby transferring signals from second messengers. S100B is also involved in cell differentiation and cell cycle progression, and it has been shown to inhibit apoptosis if applied in experimental conditions. Extracellularly, in both normal physiology and during traumatic administered conditions, S100B promotes neurogenesis and neuronal plasticity. Some authors claim that S100B is released into the serum through the disrupted the blood-brain barrier. S100B also regulates astrocyte and microglia migration and acts as a neurotrophic or neurotoxic molecule. Increased S100B concentration in blood plasma and CSF is a biochemical marker of traumatic brain injury, stroke, multiple sclerosis, neurodegenerative diseases, CNS infections and tumors [2-6].

The aim of the study was to determine and compare the S100B concentration in blood plasma and cerebrospinal fluid (CSF) patients with:

- neurodegenerative diseases including Alzheimer's disease (AD) and amyotrophic lateral sclerosis (ALS);
- disease with inflammatory and neurodegenerative etiopathogenesis – multiple sclerosis (MS);
- the reference group consisting of patients suffering from the conditions which do not alter standard parameters of CSF and blood, such as idiopathic headache and idiopathic facial nerve palsy.

# **MATERIAL AND METHODS**

## **Patients**

Blood and CSF samples were obtained from patients admitted to the Department of Neurology, Medical University of Bialystok. The study was approved by the Medical University of Bialystok Ethics Committee for Research on Humans and Animals (R-I-002/382/2009) and written consent was obtained from all subjects. All individuals underwent lumbar puncture for diagnostic purposes and MR imaging of the brain was done to exclude organic lesions such as tumors, ischemic or hemorrhagic stroke. The clinical characteristics of patients are shown in Table I.

**Table I.** The clinical characteristics of patients

	•			CSF	
Diagnosis	Number of patients (women)	Age (years)	Q(Alb)	Total protein (15 – 45 μg/mL)	Lymphocytes (N≤ 5 cell/μL)
AD	20 (13)	69.9±10.4	5.7±1.3	38.3±17.2	1.25±1
ALS	12 (9)	57.5±10.3	6.1±2.3	39,7±15,3	2.2±2.3
MS *	40 (25)	35.5±10.5	6.3±1.5	375±125	3.8±2.1
Idiopathic headache	13 (11)	40.2 ±20.4	6.5±1.3	409 ± 155	$3.0 \pm 2.1$
Idiopathic facial nerve palsy	7 (5)	48.4 ±15.3	7.5±0.6	$357 \pm 179$	$5.1 \pm 2.1$

AD - Alzheimer's disease; ALS - amyotrophic lateral sclerosis; MS - multiple sclerosis; QAlb - coefficient of albumin; \* all patients with MS in CSF presented oligoclonal bands of IgG (type 2 or 3) and mean EDSS (Expanded Disability Status Scale) score in MS group was 1.5±0.5;

The diagnosis of AD was based on NINCDS-ADRDA criteria (National Institute of Neurologic, Communicative Disorders and Stroke/Alzheimer's Disease and Related Disorders Association) [7].

All patients had progressive, significant episodic memory and cognitive functions impairment,

lasting more than six months, validated by validated by Mini Mental State Examination (MMSE) [8].

All patients with AD have been performed magnetic resonance imaging of the brain, which excluded organic lesions in the brain and showed brain atrophy. The diagnosis of relapsing-remitting MS was based on the McDonald criteria [9]. The degree of

disability of MS patients was assessed using the Expanded Disability Status Scale (EDSS), with a mean score of  $1.5 \pm 0.6$  [10]. All patients underwent head MRI study, which demonstrated multiple disseminated demyelinating plaques, with no gadolinium enhancement and a diagnostic lumbar puncture. All patients presented oligoclonal bands of IgG type 2 or 3 in CSF. None of patients was during the relapse of disease and none was treated with corticosteroids or immunomodulating drugs (beta-interferon, glatiramer acetate, natalizumab). The clinically definite ALS was diagnosed on the basis of Airlie House/El Escorial Revisited World Federation of Neurology criteria [11]. All patients with ALS showed features of damage to upper and lower motor neuron confirmed by electromyography (EMG) studies. None of patients were treated with riluzol. The reference group consisted of patients with idiopathic headache and idiopathic facial nerve palsy (Bell's palsy), who underwent lumbar puncture and CSF analysis to exclude CNS infection or subarachnoid hemorrhage. No one of patient with idiopathic facial nerve palsy was immunocompromised and had herpes simplex virus 1 antibodies in blood in ELISA tests (ELISA kit; Genzyme Virotech GmbH, Rüsselsheim, Germany).

# Samples preparation

Samples of anticoagulated blood were centrifuged and collected plasma were frozen (-80°C). After collection, the CSF underwent a standard examination. Then samples of CSF were centrifuged (2000 x g, 20 min) and supernatants were subjected to total protein analysis and frozen (-80°C). CSF analysis

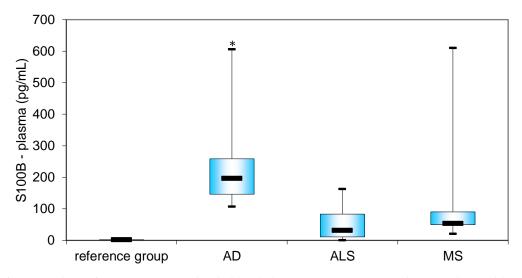
included physical properties, cytosis, total protein concentration and Q Alb ratio (Q Alb = albumin in CSF (mg) / serum albumin (g) x 1000) indicating efficiency of the blood-CSF barrier. The S100B concentration was measured with ELISA kit provided by LDN Labor Diagnostika Nord Gmbh & Co KG, Germany.

#### Statistical analysis

The differences between the groups were evaluated with an unpaired Student's t-test. The results were analyzed statistically using the Kruskal-Wallis and Dwass-Steel-Critchlow-Fligner test. The statistically significant p value was <0.05.

## RESULTS

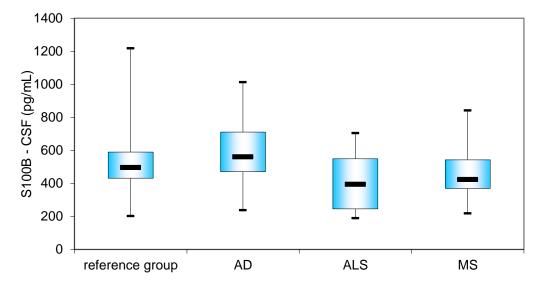
Concentrations of S100B in plasma collected from patients diagnosed with Alzheimer's disease (252,38±183,50 pg/mL) were significantly higher than in patients with amyotrophic lateral sclerosis  $(53,96\pm56,92 \text{ pg/mL})$  (p < 0,029). Concentrations of S100B in plasma collected from patients diagnosed with Alzheimer's (252,38±183,50 pg/mL) and multiple sclerosis (164,92±250,14 pg/mL) were above laboratory standards, and in patients with amyotrophic lateral sclerosis (53,96±56,92 pg/mL) and the reference group (2,12 pg/mL) were below laboratory norms included in instruction for use of S100B ELISA kit provided by LDN Labor Diagnostika Nord Gmbh & Co KG, Germany (N>75 pg/mL) (Figure 1).



 $\begin{tabular}{l} \textbf{Figure 1.} Comparison of S100B concentration in blood plasma *p < 0.029 comparing to patients with ALS; AD - Alzheimer's disease; ALS - amyotrophic lateral sclerosis; MS -multiple sclerosis; Kruskal-Wallis test with post hoc Dwass-Steele-Critchlow-Fligner test \\ \end{tabular}$ 

Concentrations of S100B in CSF collected from the reference group ( $546,96\pm236,62$  pg/mL) and from patients with Alzheimer's disease ( $587,53\pm189,57$  pg/mL), amyotrophic lateral sclerosis

(404,41±179,56 pg/mL), and multiple sclerosis (462,03±146,01 pg/mL) were very similar, and none of pairwise comparisons reached statistical significance (Figure 2).



**Figure 2.** Comparison of S100B concentration in cerebrospinal fluid (CSF); AD - Alzheimer's disease; ALS - amyotrophic lateral sclerosis; MS - multiple sclerosis; Kruskal-Wallis test with post hoc Dwass-Steele-Critchlow-Fligner test

# **DISCUSSION**

In our studies concentrations of S100B in plasma and CSF collected from patients diagnosed with AD were the highest among all evaluated diseases and in plasma were significantly higher than in patients with ALS. It is consistent with previous studies which showed highly increased concentrations of S100B in plasma and CSF of patients with AD [12,13]. AD is a neurodegenerative disease in which etiopathogenesis takes place activation and destruction of astrocytes causing S100B release. S100B triggers the formation of amyloid plaques and increases intraneuronal free calcium and nitric oxide concentration, which results in activation of inflammatory process and leads to neuronal injury. It has also been shown that betaamyloid stimulates the synthesis of S100B production in astrocytes. The most probably the high concentration of S100B in plasma of patients with AD results from its high concentration in CSF and secondary diffusion from CSF to blood due to increased permeability of the blood-CSF-barrier in the course of this disease [13,14].

The concentrations of S100B in plasma and CSF of patients with ALS are respectively decreased, which is also in agreement with previous studies. It

results from the fact that ALS is a slowly progressing neurodegenerative disease characterized, first of all, by motor neuron loss and astrogliosis with ongoing consumption and downregulation of S100B in the course of disease. S100b is also accumulated locally near damaged astrocytes and is not released into the CSF compartment [15]. In the course of ALS, the presence of S100B in blood plasma also most probably results from the alternation in blood-CSF-barrier permeability which is ongoing in the course of disease [16].

The concentrations of S100B in plasma and CSF collected from patients diagnosed with multiple sclerosis were moderately increased, which is in agreement with previous studies [17].

Neurodegeneration and inflammatory process plays an important role in the pathogenesis of MS and causes excessive CNS cell destruction with secondary release of S100B [17,18-23].

The AD, ALS and MS patients results of plasma S100B concentration were compared with laboratory norms included in instruction for use of S100B ELISA kit provided by LDN Labor Diagnostika Nord Gmbh & Co KG, Germany. There are no laboratory norms for S100B concentration in CSF that wise the obtained results of CSF S100B

concentration had to be compared with reference group. Since, there are no indications to perform lumbar puncture in healthy subjects, our reference group consisted of patients with idiopathic headache and idiopathic facial nerve palsy (Bell's palsy). The concentration of S100B in this reference group was very similar to the results of other examined groups. It may be an evidence that so called "idiopathic headaches" or "idiopathic facial nerve palsy" are not exactly idiopathic but some "organic" pathological processes are engaged in their pathogenesis. This is in agreement with previous studies which showed that S100B contributes to idiopathic headache pathology by participation in a glial activation leading to neuroinflammation [24,25].

Since S100B protein is synthesized among others by astrocytes and Schwann cells, the protein present in blood comes also from the CSF origin. Astrocytes envelop cerebral capillaries and secreted S100B can cross a damaged BBB to the blood where it is detected in plasma. Additionally, S100B is abundantly expressed by myeloid cells and adipocytes. Despite normal values of Q(Alb) ratio, the marker of the blood-CSF-barrier permeability, counted for all study groups, pathologically increased S100B protein penetration from CSF to blood due to ongoing neurodegeneration seems to be the most probable explanation of its high serum concentration in AD, ALS and MS patients. Results of our study suggest the blood-CSF-barrier is altered the most in AD patients. The S100B protein penetration in MS and especially in ALS are less affected and there were almost no protein diffusion in reference group. The AD patients were the oldest ones and a dependency of the blood-CSF-barrier permeability on age has been previously described [6,26].

# **CONCLUSIONS**

In conclusion, results of our preliminary study suggest that concentrations of S100B in plasma and CSF of patients suffering from neurodegenerative disorders may result from their pathogenesis.

Results of our studies indicate the importance of S100B protein concentration assessment in blood serum in CNS diseases differential diagnostics.

# **Conflict of interests**

None declared.

## Financial disclosure

None declared.

# **REFERENCES**

- Steiner J, Bernstein HG, Bielau H, Berndt A, Brisch R, Mawrin C, Keilhoff G, Bogerts B. Evidence for a wide extra-astrocytic distribution of S100B in human brain. BMC Neurosci. 2007 Jan 2:8:2.
- Steiner J, Bernstein HG, Bogerts B, Gos T, Richter-Landsberg C, Wunderlich MT, Keilhoff G. S100B is expressed in, and released from, OLN-93 oligodendrocytes: Influence of serum and glucose deprivation. Neuroscience. 2008 Jun 23;154(2):496-503.
- 3. Ondruschka B, Pohlers D, Sommer G, Schober K, Teupser D, Franke H, Dressler J. S100B and NSE as useful postmortem biochemical markers of traumatic brain injury in autopsy cases. J Neurotrauma. 2013 Nov 15;30(22):1862-71.
- Park JW, Suh GI, Shin HE. Association between cerebrospinal fluid S100B protein and neuronal damage in patients with central nervous system infections. Yonsei Med J. 2013 May 1;54(3):567-71
- Sorci G, Bianchi R, Riuzzi F, Tubaro C, Arcuri C, Giambanco I, Donato R. S100B Protein, A Damage-Associated Molecular Pattern Protein in the Brain and Heart, and Beyond. Cardiovasc Psychiatry Neurol. 2010;2010.
- 6. Yardan T, Erenler AK, Baydin A, Aydin K, Cokluk C. Usefulness of S100B protein in neurological disorders. J Pak Med Assoc. 2011 Mar;61(3):276-81.
- 7. Dubois B, Feldman HH, Jacova C, Dekosky ST, Barberger-Gateau P, Cummings J, Delacourte A, Galasko D, Gauthier S, Jicha G, Meguro K, O'brien J, Pasquier F, Robert P, Rossor M, Salloway S, Stern Y, Visser PJ, Scheltens P. Research criteria for the diagnosis of Alzheimer's disease: revising the NINCDS-ADRDA criteria. Lancet Neurol. 2007 Aug;6(8):734-46.
- 8. Pangman VC, Sloan J, Guse L. An examination of psychometric properties of the mini-mental state examination and the standardized minimental state examination: implications for clinical practice. Appl Nurs Res. 2000 Nov;13(4):209-13.
- Polman CH, Reingold SC, Banwell B, Clanet M, Cohen JA, Filippi M, Fujihara K, Havrdova E, Hutchinson M, Kappos L, Lublin FD, Montalban X, O'Connor P, Sandberg-Wollheim M, Thompson AJ, Waubant E, Weinshenker B, Wolinsky JS. Diagnostic criteria for multiple sclerosis: 2010 revisions to the McDonald criteria. Ann Neurol. 2011 Feb;69(2):292-302.
- 10. Kurtzke JF. Rating neurologic impairment in multiple sclerosis: an expanded disability status

- scale (EDSS). Neurology. 1983 Nov;33(11): 1444-52.
- 11. Brooks BR, Miller RG, Swash M, Munsat TL, Diseases WFoNRGoMN. El Escorial revisited: revised criteria for the diagnosis of amyotrophic lateral sclerosis. Amyotroph Lateral Scler Other Motor Neuron Disord. 2000 Dec;1(5):293-9.
- 12. Peskind ER, Griffin WS, Akama KT, Raskind MA, Van Eldik LJ. Cerebrospinal fluid S100B is elevated in the earlier stages of Alzheimer's disease. Neurochem Int. 2001 Nov-Dec;39(5-6):409-13.
- Chaves ML, Camozzato AL, Ferreira ED, Piazenski I, Kochhann R, Dall'Igna O, Mazzini GS, Souza DO, Portela LV. Serum levels of S100B and NSE proteins in Alzheimer's disease patients. J Neuroinflammation. 2010 Jan 27;7:6.
- Mrak RE, Sheng JG, Griffin WS. Glial cytokines in Alzheimer's disease: review and pathogenic implications. Hum Pathol. 1995 Aug;26(8):816-23.
- 15. Süssmuth SD, Tumani H, Ecker D, Ludolph AC. Amyotrophic lateral sclerosis: disease stage related changes of tau protein and S100 beta in cerebrospinal fluid and creatine kinase in serum. Neurosci Lett. 2003 Dec 15;353(1):57-60.
- 16. Garbuzova-Davis S, Sanberg PR. Blood-CNS Barrier Impairment in ALS patients versus an animal model. Front Cell Neurosci. 2014 Feb 3:8:21.
- 17. Dutta R, Trapp BD. Mechanisms of neuronal dysfunction and degeneration in multiple sclerosis. Prog Neurobiol. 2011 Jan;93(1):1-12.
- 18. Barateiro A, Afonso V, Santos G, Cerqueira JJ, Brites D, van Horssen J, Fernandes A. S100B as a Potential Biomarker and Therapeutic Target in Multiple Sclerosis. Mol Neurobiol. 2016 Aug;53(6):3976-3991.
- 19. Bennett JL, Stüve O. Update on inflammation, neurodegeneration, and immunoregulation in

- multiple sclerosis: therapeutic implications. Clin Neuropharmacol. 2009 May-Jun;32(3):121-32.
- 20. Hohlfeld R. Biotechnological agents for the immunotherapy of multiple sclerosis. Principles, problems and perspectives. Brain. 1997 May;120 (Pt 5):865-916.
- Micu I, Jiang Q, Coderre E, Ridsdale A, Zhang L, Woulfe J, Yin X, Trapp BD, McRory JE, Rehak R, Zamponi GW, Wang W, Stys PK. NMDA receptors mediate calcium accumulation in myelin during chemical ischaemia. Nature. 2006 Feb 23:439(7079):988-92.
- 22. Nave KA, Trapp BD. Axon-glial signaling and the glial support of axon function. Annu Rev Neurosci. 2008;31:535-61.
- 23. Trapp BD, Nave KA. Multiple sclerosis: an immune or neurodegenerative disorder? Annu Rev Neurosci. 2008;31:247-69.
- 24. Teepker M, Munk K, Mylius V, Haag A, Möller JC, Oertel WH, Schepelmann K. Serum concentrations of s100b and NSE in migraine. Headache. 2009 Feb;49(2):245-52.
- 25. Papandreou O, Soldatou A, Tsitsika A, Kariyannis C, Papandreou T, Zachariadi A, Papassotiriou I, Chrousos GP. Serum S100beta protein in children with acute recurrent headache: a potentially useful marker for migraine. Headache. 2005 Nov-Dec;45(10):1313-6.
- 26. Akiyama H, Barger S, Barnum S, Bradt B, Bauer J, Cole GM, Cooper NR, Eikelenboom P, Emmerling M, Fiebich BL, Finch CE, Frautschy S, Griffin WS, Hampel H, Hull M, Landreth G, Lue L, Mrak R, Mackenzie IR, McGeer PL, O'Banion MK, Pachter J, Pasinetti G, Plata-Salaman C, Rogers J, Rydel R, Shen Y, Streit W, Strohmeyer R, Tooyoma I, Van Muiswinkel FL, Veerhuis R, Walker D, Webster S, Wegrzyniak B, Wenk G, Wyss-Coray T. Inflammation and Alzheimer's disease. Neurobiol Aging. 2000;21(3):383-421.